

# UC Irvine

## UC Irvine Previously Published Works

### Title

Dorsoventral differences in intrinsic properties in developing CA1 pyramidal cells.

### Permalink

<https://escholarship.org/uc/item/94m6g9fp>

### Journal

The Journal of neuroscience : the official journal of the Society for Neuroscience, 32(11)

### ISSN

0270-6474

### Authors

Marcelin, Béatrice  
Liu, Zhiqiang  
Chen, Yuncai  
et al.

### Publication Date

2012-03-01

### DOI

10.1523/jneurosci.5870-11.2012

### Copyright Information

This work is made available under the terms of a Creative Commons Attribution License, available at <https://creativecommons.org/licenses/by/4.0/>

Peer reviewed

Published in final edited form as:

*J Neurosci.* 2012 March 14; 32(11): 3736–3747. doi:10.1523/JNEUROSCI.5870-11.2012.

## Dorsal-ventral differences in intrinsic properties in developing CA1 pyramidal cells

Béatrice Marcelin<sup>1,2,\*</sup>, Zhiqiang Liu<sup>3,\*</sup>, Yuncai Chen<sup>4,\*</sup>, Alan S. Lewis<sup>3</sup>, Albert Becker<sup>5</sup>, Shawn McClelland<sup>4</sup>, Dane M. Chetkovich<sup>3,6</sup>, Michele Migliore<sup>7</sup>, Tallie Z. Baram<sup>4</sup>, Monique Esclapez<sup>1,2</sup>, and Christophe Bernard<sup>1,2</sup>

<sup>1</sup>INSERM, U1106, Marseille, F-13385, France

<sup>2</sup>Aix Marseille Université, Faculté de Médecine, Marseille, F-13385, France

<sup>3</sup>Davee Department of Neurology and Clinical Neurosciences, Feinberg School of Medicine, Northwestern University, Chicago, Illinois 60611, USA

<sup>4</sup>Departments of Anatomy/Neurobiology and Pediatrics, University of California-Irvine Irvine CA 92697-4475, USA

<sup>5</sup>Department of Neuropathology, University of Bonn Medical Center, Sigmund Freud Strasse 25, 53105 Bonn, Germany

<sup>6</sup>Department of Physiology, Feinberg School of Medicine, Northwestern University, Chicago, Illinois 60611, USA

<sup>7</sup>Institute of Biophysics, National Research Council, Palermo, Italy

### Abstract

The dorso-ventral and developmental gradients of entorhinal layer II cell grid properties correlate with their resonance properties and with their hyperpolarization-activated cyclic nucleotide-gated ion channel current (HCN,  $I_h$ ) characteristics. We investigated whether such correlation existed in rat hippocampal CA1 pyramidal cells, where place fields also show spatial and temporal gradients. Resonance was absent during the first postnatal week, and emerged during the second week. Resonance was stronger in dorsal than ventral cells, in accord with HCN current properties. Resonance responded to cAMP in ventral but not in dorsal cells. The dorso-ventral distribution of HCN1 and HCN2 subunits and of the auxiliary protein tetratricopeptide repeat-containing Rab8b-interacting protein (TRIP8b) could account for these differences between dorsal and ventral cells. The analogous distribution of the intrinsic properties of entorhinal stellate and hippocampal cells suggests the existence of general rules of organization among structures that process complementary features of the environment.

### Introduction

The rodent hippocampus is not a homogenous structure (Moser and Moser, 1998). It can be divided into three compartments (dorsal, intermediate and ventral), each with specific anatomical features (Swanson et al., 1978; Fanselow and Dong, 2010). Gene expression analysis reveals that the dorsal and ventral regions of the hippocampus have distinct

Corresponding author: Christophe Bernard, INSERM, U751, Faculté de Médecine, 27 Bd Jean Moulin, Marseille, F-13385, France, christophe.bernard@univmed.fr.

\*equally contributing authors

Conflict of interest: none.

molecular organizations (Thompson et al., 2008; Dong et al., 2009). This heterogeneity likely underlies their distinct functions; the dorsal (septal) part dealing with cognitive processes (memory) and the ventral (temporal) part dealing with emotional aspects (Fanselow and Dong, 2010). At the cellular level, place fields coded by CA1 pyramidal cells (O'Keefe and Nadel, 1978) expand along the dorso-ventral axis and their properties change during development (O'Keefe and Nadel, 1978; Kjelstrup et al., 2008; Langston et al., 2010; Wills et al., 2010). Given the different information processing performed in the dorsal and ventral parts, we hypothesized that hippocampal neurons would have different integrative properties along the dorso-ventral axis.

Pioneering work performed in the entorhinal cortex provides ground for such a hypothesis. Layer II stellate cells represent the external space as a grid, with size that expands along the dorso-ventral axis and with properties that change during development (Sargolini et al., 2006; Langston et al., 2010; Wills et al., 2010). In parallel, the integrative properties of layer II stellate cells change in time (during development) and space (along the dorso-ventral axis), and these correlate with differential distributions of HCN channels and leak K conductances in these neurons, (Nolan et al., 2007; Giocomo et al., 2007; Garden et al., 2008; Giocomo and Hasselmo, 2008; Burton et al., 2008; Boehlen et al., 2010; Yoshida et al., 2011).

Hippocampal CA1 pyramidal cells are optimized to preferentially respond, or resonate, to inputs in the theta frequency range, similar to stellate cells. Resonance is achieved by specific ion channels, including the hyperpolarization-activated current  $I_h$  (Hutcheon and Yarom, 2000; Pike et al., 2000; Hu et al., 2002; Narayanan and Johnston, 2007; Marcelin et al., 2009). HCN channels also play a key role in temporal coding. They provide negative time delays in the theta frequency range (Narayanan and Johnston, 2008; Marcelin et al., 2009). They also shape synaptic inputs in the gamma (40–80 Hz) band (Magee, 1998; Garden et al., 2008). The developmental profile of the expression of HCN channel isoforms and of  $I_h$  properties in the CA1 region (Bender et al., 2001; Vasilyev and Barish, 2002; Surges et al., 2006; Brewster et al., 2007), suggests that integrative properties of pyramidal cells will be age-dependent.

Although not in direct synaptic contact, layer II grid cells and hippocampal place cells are part of a network representing space. Therefore, we sought to study whether the spatial and developmental profiles of intrinsic properties in CA1 cells follow the same general rules as layer II stellate cells (Nolan et al., 2007; Giocomo et al., 2007; Garden et al., 2008; Giocomo and Hasselmo, 2008; Burton et al., 2008; Boehlen et al., 2010); focusing on theta resonance, temporal summation and HCN current properties.

## Materials and Methods

Hippocampal slices from the dorsal (coronal sectioning) and the ventral (with a 30° angle from the sagittal plane as in (Bernard et al., 2004)) hippocampus were prepared from postnatal (PN) day 11–12, 14–15 and PN 5 male wistar rats. ACSF contained (in mM) NaCl 126, KCl 3.5, CaCl<sub>2</sub> 2, MgCl<sub>2</sub> 1.3, NaH<sub>2</sub>PO<sub>4</sub> 1.2, NaHCO<sub>3</sub> 26, D-Glucose 10, and NBQX (1 μM), D-APV (50 μM) and bicuculline (10 μM) to block AMPA, NMDA and GABA<sub>A</sub> receptors, respectively. Cells were recorded at 34 ± 1°C with a solution containing (in mM) KMeSO<sub>4</sub> 120, KCl 20, EGTA 0.2, MgCl<sub>2</sub> 2, HEPES 10, Na<sub>2</sub>ATP 4, Tris GTP 0.3, Phosphocreatine 14, biocytin 0.4% and KOH to adjust to pH 7.3 (whole cell recordings); or KCl 120, tetraethylammonium-Cl 20, HEPES 10, 4-aminopyridine 5, CaCl<sub>2</sub> 2, MgCl<sub>2</sub> 1, BaCl<sub>2</sub> 1, biocytin 0.4% and KOH to adjust to pH 7.3 (cell attached recordings). In some experiments, 10 μM cAMP was added to the pipette solution. Membrane potential was not corrected for liquid junction potential (2 mV, cell attached; 9 mV, whole cell). For

consistency, all recordings were performed in mid-distal CA1 along the transverse axis, close to the stratum oriens border for somata, and close to stratum lacunosum moleculare for dendrites.

CA1 pyramidal cells somata and dendrites were recorded under visual control (Nikon FN1 microscope – Scientifica Patch Star manipulators) with an Axopatch700B amplifier and Digidata 1322 interface (Axon Instruments). The distance between the recording site and the soma was determined both under visual control before performing dendritic recordings, and *post hoc* following morphological processing as previously described (Esclapez et al., 1999).

### Analysis of $I_h$ (whole cell recordings)

Currents mediated by  $I_h$  were recorded in voltage-clamp mode by applying hyperpolarizing voltage steps (starting from a holding potential of  $-50$  mV up to  $-140$  mV). Data were acquired with the help of pCLAMP 10 and analysed offline using Clampfit 10.0 (Axon Instruments). The amplitude of  $I_h$  was determined by subtracting the instantaneous current at the beginning of the voltage step from the steady-state current at the end. The activation time constant was obtained using a double exponential. Boltzmann fits of the tail current were made to obtain  $V_{1/2}$ , the midpoint activation voltage. Single or double exponential fits were used to measure the time constants of  $I_h$  activation. In a subset of experiments,  $I_h$  properties were obtained by subtracting the current traces before and after the application of  $10$   $\mu$ M of the  $I_h$  blocker ZD7288.

### Analysis of $I_h$ (cell attached recordings)

Data collection started after obtaining  $> 2$  G $\Omega$  seal. We used  $6$  M $\Omega$  pipette resistance for somatic recordings and  $12$  M $\Omega$  for dendritic recordings. A family of currents was obtained from  $+10$  to  $-90$  mV from rest (effective change in trans-membrane potential of the patch), in  $10$  mV increments. A series of 4 leak currents ( $+30$  mV from rest) was obtained and averaged. The averaged leak current was subtracted from the family of currents (the leak current was multiplied by the ratio of test voltage command/leak voltage command). This operation gave the activation curve of  $I_h$ . The whole procedure was repeated 10 times, and the activation curves were then averaged. At the end of the experiment, a gentle suction was applied to break-in in order to measure the resting membrane potential.  $I_h$  parameters were measured as described above for whole cell recordings.

### Resonance protocol

Discrete current wavelets at varying frequencies were injected in the recorded cell in current-clamp mode (from  $1$  to  $50$  Hz, with  $1$  Hz steps; and 25 wavelets between  $50$  and  $200$  Hz with logarithmic scale spacing); each wavelet consisted of 12 cycles with waxing and waning amplitude. The amplitude of the injected current was adjusted so that the  $1$  Hz wavelet produced a  $\pm 5$  mV excursion of membrane potential. A  $50$  ms hyperpolarizing step was applied between two successive wavelets. We used discrete wavelet injections rather than the common ZAP/Chirp stimulus, as described (Marcelin et al., 2009). Impedance ( $Z$ ) was calculated as the ratio of the voltage/current Fourier transforms:  $Z = \text{FFT}(\text{voltage response})/\text{FFT}(\text{injected current})$ , for each frequency. The magnitude of the complex-valued impedance  $|Z|$  was plotted against frequency to give an impedance–magnitude profile. Two resonance parameters were measured:  $F_{\text{res}}$ , the resonance frequency when  $|Z|$  is maximum; and  $Q$ , the amplification ratio defined as the  $|Z|_{F_{\text{res}}}/|Z|_{1 \text{ Hz}}$  ratio. The impedance phase was calculated as  $\varphi = \tan^{-1}(\text{Im}(Z)/\text{Re}(Z))$ , where  $\text{Im}(Z)$  and  $\text{Re}(Z)$  are the imaginary and real parts of  $Z$ , respectively (Narayanan and Johnston, 2008).  $F_{\varphi}$  is defined as the frequency  $f$  where  $\varphi(f) = 0$ . For  $f < F_{\varphi}$ ,  $\varphi(f) > 0$  and the response of the membrane appears to occur before the injected current (phase lead). Conversely, for  $f > F_{\varphi}$ ,  $\varphi(f) < 0$  and the response of the membrane appears to occur after the injected current (phase lag). To characterize the

amount of phase lead, we measured  $\Phi_L$ , the total inductive phase, defined as the area of  $\varphi(f)$  where  $\varphi(f) > 0$  (Narayanan and Johnston, 2008).

### Temporal summation

Artificial excitatory postsynaptic potentials (EPSPs) were evoked by current injections of the form of an alpha function:  $I = I_{\max} t \exp(-\alpha t)$ , with  $\alpha = 0.1$ . Temporal summation ratio in a train of 5 artificial EPSPs was computed as EPSP5/EPSP1, where EPSP5 and EPSP1 are the amplitudes of first and fifth artificial EPSPs in the train, respectively.

### Morphology

After electrophysiological recordings, slices were fixed overnight at 4°C in a solution containing 4% paraformaldehyde in 0.1 M phosphate buffer (PB; pH 7.4). After fixation, slices were rinsed in PB, cryoprotected in sucrose, and quickly frozen on dry ice. Detection of labeled neurons was performed on unsectioned slices as described previously (Esclapez et al., 1999). Briefly, to neutralize endogenous peroxidase, slices were pretreated for 30 minutes in 1% H<sub>2</sub>O<sub>2</sub>. After several rinses in 0.1M phosphate buffered saline (pH 7.4; PBS), slices were incubated overnight at 4°C in avidin-biotinylated peroxidase complex (1:200; Vector Laboratories, Burlingame, CA) diluted in PBS. After 1.5 hr rinses in PBS, slices were processed with 3,3'-diaminobenzidine tetrahydrochloride (DAB; Sigma) and H<sub>2</sub>O<sub>2</sub> diluted in distilled H<sub>2</sub>O (Sigma fast tablets). A morphometric analysis of pyramidal cells was performed after three-dimensional reconstruction using a Neurolucida workstation. The morphometric data did not involve compensation for tissue shrinkage, which was 86 % in z and 10 % in x/y axes.

### Simulations

The Neurolucida morphometric data of 10 CA1 cells (5 from dorsal and 5 from ventral region) was imported into the NEURON simulation environment (v7.1, (Hines and Carnevale, 1997)), and a multicompartmental model for each of them was implemented using the same passive properties ( $\tau_m = 28\text{ms}$ ,  $R_m = 28\text{k}\Omega \cdot \text{cm}^2$ ,  $R_a = 150\Omega \cdot \text{cm}$ ) in all cases. The NEURON Impedance Tools was then used to calculate the voltage attenuation measured at the soma when the same current is injected in every point on the cell.

### Western blotting

Tissue preparation and western blotting were performed as previously described (Lewis et al., 2009) with some modifications. Briefly, animals were euthanized and hippocampi were rapidly removed and divided into three parts along the septo-temporal axis: dorsal, intermediate and ventral parts. The samples were immediately frozen in liquid nitrogen and stored at -80°C. For the preparation of tissue homogenate, the hippocampal samples were homogenized in lysis buffer containing 50 mM Tris-HCl (pH 7.4), 150 mM NaCl, 1% Nonidet-40, 0.1% sodium dodecyl sulfate, 0.5% sodium desoxycholate, 5 mM EDTA, and a mixture of protease inhibitors (Roche). The homogenates were centrifuged at 20,000g at 4°C for 10 min, and the supernatant was collected and frozen until further use. Protein concentrations were determined with the Bradford protein assay (Bio-Rad). For Western blotting, extracts were mixed with sample buffer, boiled at 95°C for 5 min, cooled on ice for 2 min and separated on 10% SDS-PAGE at 150 volts under denaturing conditions. Proteins were blotted on PVDF membranes and the blots were blocked in 5% milk in TBST solution for 30 min, and incubated with primary antibodies at 4°C overnight. Then the blots were rinsed in TBST for 3×10 min and incubated in HRP-conjugated secondary antibody for 1 hour. Antibody binding was detected using Super Signal West Pico Chemiluminescent Substrate (Thermo), and quantified by densitometry from blots with linear signal using NIH Image J software.  $\alpha$ -tubulin expression was used for loading control. The relative

proportions were calculated by dividing the value of  $\alpha$ -tubulin and normalized ventral density by dorsal. The following antibodies and dilutions were used for western blotting in the present study: guinea pig anti-HCN1 (Shin and Chetkovich, 2007) 1:1,000; guinea pig anti-HCN2 (Shin et al., 2006) 1:1,000; rabbit anti-TRIP8b (Lewis et al., 2009) 1:5,000; mouse anti- $\alpha$ -tubulin (DM1A, Santa Cruz, CA) 1:5,000. Normalized data were analyzed using unpaired, two-tailed Student's t-test, with significance considered as  $p < 0.05$ .

### mRNA isolation of hippocampi and real-time RT-PCR

Samples were prepared and stored at  $-80^{\circ}\text{C}$  as above. mRNA from the dorsal and ventral parts of the hippocampus was isolated by Dynabeads mRNA Direct Micro Kit (Dynal, Oslo) according to the manufacturer's protocol. HCN1 and HCN2 transcript quantification was performed by real-time RT-PCR (PE Biosystems PRISM 7700, Foster City). Primers and double-fluorescent oligonucleotides (FAM=reporter dye, TAMRA=quencher dye) for HCN1 and HCN2 channel subunits and synaptophysin were designed with Primer Express software (PE Biosystems, Foster City). No significant homology of the amplicon sequences with other previously characterized genes has been found searching GenBank data bases by BLASTN program. Relative quantification of the starting mRNA copy numbers using multiple replicates for each reaction was performed according to the  $\Delta\Delta\text{Ct}$ -method (Fink et al., 1998). The signal threshold was set within the exponential phase of the reaction for determination of the threshold cycle (Ct). Synaptophysin was used as endogenous, neuron-specific reference gene for normalization of HCN channel subunit mRNAs. Relative quantification started from 5.0 – 10.0 ng of mRNA. RT-PCR reactions were carried out in a total reaction volume of 12.5  $\mu\text{l}$  using the EZ RT-PCR Kit (PE Biosystems, Foster City). Reactions started with 300  $\mu\text{M}$  of each dNTP, 1xTaqMan EZ Buffer, 0.1U/ $\mu\text{l}$  rTth DNA polymerase, 0.01U/ $\mu\text{l}$  AmpErase UNG, Mn(OAc)<sub>2</sub> and primers as well as double-fluorescent oligonucleotides. Cycling conditions included  $50^{\circ}\text{C}$  for 2min,  $60^{\circ}\text{C}$  for 20min (RT reaction), followed by  $95^{\circ}\text{C}$  for 5min, and a two-step PCR with 60 cycles at  $94^{\circ}\text{C}$  for 15sec and  $59^{\circ}\text{C}$  for 60sec (PE Biosystems PRISM 7700 SDS, Foster City). Fluorescent signals were normalized to a passive internal reference ( $\Delta\text{Rn}$ ) and the threshold cycle (Ct) was set within cycle 28, i.e. the exponential phase of the PCR. Based on the Ct ratios, the starting copy number of HCN1 and HCN2 mRNAs were calculated relative to the reference gene ( $\Delta\Delta\text{Ct}$  method;  $n=5$  animals for each group and each time point). The primer sequences for PCR reactions can be found in (Jung et al., 2011). Group comparisons were carried out by a Mann-Whitney-U-test.

### Tissue preparation and immunocytochemistry (ICC) of brain slices

Sprague-Dawley rats aged 12 days ( $n = 4$ ) were perfused via the ascending aorta with 0.9% saline solution followed by perfusion with 4% paraformaldehyde solution made in 0.1 M PB (pH 7.4,  $4^{\circ}\text{C}$ ). Brains were postfixed for 2–4 hrs ( $4^{\circ}\text{C}$ ) and immersed in 15%, followed by 25% sucrose for cryoprotection. Serial sections (20  $\mu\text{m}$  thickness) were first collected from the septal pole of the hippocampus (cut in the coronal plane). Brain blocks were then turned to permit collection of horizontal sections from the temporal pole (Bender et al., 2005). In each plane, 1 in 6 serial sections were subjected to HCN1-ICC and an adjacent series of sections was used for MAP2-ICC. ICC was performed on free-floating sections using standard avidin-biotin complex methods, as described previously (Chen et al., 2001; Brewster et al., 2007; Bender et al., 2007). Briefly, after several washes with PBS containing 0.3% Triton X-100 (PBS-T, pH 7.4), sections were treated with 0.3% H<sub>2</sub>O<sub>2</sub>/PBS for 30 min, then blocked with 5% normal goat serum for 30 min in order to prevent non-specific binding. After rinsing, sections were incubated for 36 hrs at  $4^{\circ}\text{C}$  with guinea pig anti-HCN1 (1:5,000; a gift from Dr. Shigemoto) or mouse anti-MAP2 (1:50,000, Sigma) antisera in PBS containing 1% BSA, and washed in PBS-T ( $3 \times 5$  minutes). Sections were incubated with biotinylated goat-anti-guinea pig or mouse IgG (1:400, Vector laboratories,



Burlingame, CA) in PBS for 1 hr at room temperature. After washing ( $3 \times 5$  min), sections were incubated with the avidin-biotin-peroxidase complex solution (1:200, Vector) for 2 hrs, rinsed ( $3 \times 5$  min), and reacted with 0.04% 3,3'-diaminobenzidine (DAB) containing 0.01% H<sub>2</sub>O<sub>2</sub>.

## Chemicals

All chemicals and drugs were obtained from Sigma or Ascent unless specified otherwise.

We used non parametric Mann-Whitney test for  $n < 6$ , and Student's t-test or ANOVA otherwise for electrophysiological data and non-parametric two independent Sample Wilcoxon Rank Sum test for the morphometric analysis. Unless noted otherwise, results are expressed as mean  $\pm$  s.e.m, with  $p < 0.05$  considered significant.

## Results

### Morphological properties

The hippocampus develops in a dorso-ventral gradient (Bayer, 1980). Since most recordings were performed at PN11-12, we first investigated the morphological features of CA1 pyramidal cells from the dorsal and ventral parts of the hippocampus (Fig. 1). Morphometric analysis revealed no significant differences in mean cell body area, mean total length of apical and basal dendrites, as well as in the mean diameter of the 1<sup>st</sup> order apical dendrite between CA1 pyramidal cells from the dorsal ( $n=5$ ) and ventral ( $n=5$ ) hippocampus at PN11-12 (Table 1). To test whether neurons from the dorsal and ventral regions have similar signal propagation features, we implemented a multicompartamental model from 3D reconstructions of the cells shown in Figure 1. Using the same passive properties for all model neurons, the average input resistances measured from the soma were  $239.7 \pm 49.3$  M $\Omega$  (dorsal) and  $178.7 \pm 26$  M $\Omega$  (ventral), not significantly different (Mann-Whitney Rank Sum Test,  $p=0.421$ ). As shown in Fig. 2, the voltage attenuation measured at the soma was also comparable at all distances, suggesting that the expected overall signal propagation in these neurons would be relatively similar. We conclude that the neurons recorded in this study had similar morphological and passive cable properties.

### Theta resonance and phase responses are larger in dorsal than in ventral cells

With the use of whole-cell recordings, we first assessed the resonance properties of hippocampal CA1 pyramidal cell somata. We measured the resonance frequency ( $F_{res}$ ), the optimal frequency giving rise to the largest response, and the amplification factor  $Q$  at  $F_{res}$  (compared to 1 Hz). Injection of oscillating currents resulted in larger resonance in dorsal ( $n=46$ ) than in ventral somata ( $n=40$ ) at PN11-12 (Table 2, Fig. 3A–B). Resonance depended upon  $I_h$  because it was blocked by the  $I_h$  antagonist ZD7288 (10  $\mu$ M) in dorsal ( $n=9$ ) and ventral ( $n=8$ ) cells (Fig. 3A). Impedance curves were identical in dorsal and ventral cells in the presence of ZD7288 (Fig. 3C), suggesting that the dorso-ventral differences in resonance were due to  $I_h$ .

HCN channels not only provide band-pass filtering properties, directly controlling the amplitude of the membrane response, but also impact the temporal response (Narayanan and Johnston, 2007; Narayanan and Johnston, 2008; Marcelin et al., 2009). They create an apparent negative time delay up to a critical frequency  $F_\phi$ , i.e. the voltage response of the membrane appears to precede the inputs generating this response (phase lead). Beyond  $F_\phi$ , the voltage response follows the input (phase lag). We thus evaluated the phase response  $\phi(f)$  as a function of the frequency of the input current in CA1 pyramidal cell somata. We determined  $F_\phi$  the frequency where  $\phi(f) = 0$  (i.e. where the transition from phase lead to phase lag occurs), and  $\Phi_L$ , the total inductive phase, a measure of the extent of phase lead

(Fig. 3D). Phase lag and inductive phase were stronger in dorsal (n=46) than in ventral somata (n=40) at PN11-12 (Table 2, Fig. 3D–E). Phase lead depended upon  $I_h$  because it was abolished by ZD7288 (10  $\mu$ M) in dorsal (n=9) and ventral (n=8) cells (Fig. 3D). Phase curves were identical in dorsal and ventral cells in the presence of ZD7288 (Fig. 3F), suggesting that the dorso-ventral differences in phase responses were due to  $I_h$ .

Since resonance and phase lead were already present at PN11-12, we assessed whether they were also present one week earlier. Resonance and phase lead were absent at PN5 in dorsal somata (Fres =  $1.5 \pm 0.2$  Hz, Q =  $1.02 \pm 0.01$ , RMP =  $-57.8 \pm 2.4$  mV, Rin =  $642 \pm 103$  M $\Omega$ , C =  $71 \pm 5$  pF, n = 7, not shown).

### Increased temporal summation in ventral cells

Temporal summation of synaptic inputs plays a key role in information processing. Temporal summation results from a complex interplay between synaptic currents and ionic channels. Using injections of artificial synaptic currents through the recording pipette, we found that temporal summation was larger in ventral than dorsal cells (Fig. 4A–B; at 25 Hz, dorsal  $-9.4 \pm 2.5\%$ , n=10; ventral  $5.6 \pm 4.5\%$ , n=10,  $p < 0.05$ ; at 50 Hz, dorsal  $52 \pm 8\%$ , n=10; ventral  $76 \pm 8\%$ , n=10,  $p < 0.05$ ). This integrative property depended upon  $I_h$  (Magee, 1998), as temporal summation was significantly increased and the differences between dorsal and ventral cells abolished in the presence of ZD7288 (Fig. 4A–B; at 25 Hz, dorsal  $38 \pm 7\%$ , n=10; ventral  $47 \pm 8\%$ , n=10; at 50 Hz, dorsal  $137 \pm 9\%$ , n=10; ventral  $125 \pm 7\%$ , n=10,  $p > 0.7$  for both).

### Larger $I_h$ amplitude and faster kinetics in dorsal than ventral CA1 pyramidal cell somata

The dorso-ventral gradient of HCN current properties constitutes another correlate of the gradient of grid size in stellate cells. In adult neurons, in the range of potential tested here, theta resonance depends partly upon  $I_h$  availability (Narayanan and Johnston, 2007; Marcelin et al., 2009). We thus measured  $I_h$  parameters in the same cells in which we characterized resonance and temporal summation. The amplitude of  $I_h$  was larger in dorsal than ventral somata (Table 2; Fig. 5A–B). The membrane potential for half activation ( $V_{1/2}$ ) was more depolarized by 2.4 mV in dorsal as compared to ventral somata (Table 2; Fig. 5B).  $I_h$  activated faster in dorsal than ventral somata, whilst the slow time constant was not different (Table 2; Fig. 5B;  $p = 0.39$ ). Since  $I_h$  and resonance were measured in each recorded cell, we could correlate both parameters. As previously reported in adult ventral cells (Marcelin et al., 2009), there was a linear relationship between  $I_h$  amplitude and the amplification ratio Q (Fig. 5C).

In keeping with the increased availability of  $I_h$ , resting membrane potential was more depolarized and input resistance smaller in dorsal than ventral cells (Table 2). Input resistance was equalized in the presence of ZD7288 (dorsal:  $264 \pm 15$  M $\Omega$ , n=15; ventral:  $261 \pm 18$  M $\Omega$ , n=14;  $p = 0.92$ ). Similarly, the resting membrane potential was equalized in the presence of ZD7288 (dorsal:  $-68.0 \pm 0.8$  mV, n=15; ventral:  $-67.2 \pm 0.8$  mV, n=14;  $p = 0.51$ ).

Consistent with the minimal/absent resonance, at PN5, the amplitude of  $I_h$  was small and with slow kinetics in somata (n=7, at  $-140$  mV,  $120 \pm 20$  pA; at  $-90$  mV,  $31 \pm 2$  pA;  $V_{1/2} = -103.3 \pm 2.7$  mV;  $\tau_{\text{fast}} = 74 \pm 6$  ms;  $\tau_{\text{slow}} = 402 \pm 48$  ms).

### Identical resonance/phase responses and $I_h$ properties in proximal dendrites of dorsal and ventral cells

Resonance shows a somato-dendritic gradient in adult CA1 pyramidal cells (Narayanan and Johnston, 2007; Marcelin et al., 2009). Dendritic recordings were attempted at PN 11–12,



the earliest time point when it was technically feasible far away enough (around 130  $\mu\text{m}$ , i.e. in stratum radiatum) from the soma (Table 2). Resonance and phase responses were similar in dorsal ( $n=19$ ) and ventral ( $n=15$ ) dendrites (Table 2, Fig. 3B, ANOVA  $p=0.59$  for  $F_{\text{res}}$ ,  $p=0.79$  for  $Q$ ,  $p>0.6$  for both  $F_{\phi}$  and  $\Phi_L$ ). Resonance was statistically larger in somata than in dendrites of dorsal cells (Fig. 1B), but similar in ventral cells (ANOVA for  $F_{\text{res}}$ ,  $p=0.43$ ; for  $Q$ ,  $p=0.91$ ). In dendrites,  $I_h$  properties were similar in the ventral and dorsal cells (Table 2; Fig. 5B; ANOVA  $p>0.22$  for all parameters).

### Properties of HCN channels in somata and dendrites

In the previous sections, we characterized  $I_h$  properties using whole cell recordings because we wanted to measure both  $I_h$  and resonance in the same cell enabling their correlation (Fig. 5C). However, whole cell recordings provide only a rough estimate of channel distribution and properties. We thus performed cell attached recordings of CA1 pyramidal cell somata and dendrites at PN11-12.  $I_h$  current density was larger and  $V_{1/2}$  more depolarized in dorsal ( $n=8$ ) than ventral ( $n=8$ ) somata (Table 3, Fig. 6,  $3.8 \pm 0.5 \text{ pA}/\mu\text{m}^2$  vs.  $1.9 \pm 0.5 \text{ pA}/\mu\text{m}^2$ ,  $p<0.015$ ;  $V_{1/2} = -99.2 \pm 1.8 \text{ mV}$  vs.  $V_{1/2} = -109.9 \pm 2.7 \text{ mV}$ ,  $p<0.003$ ). In proximal dendrites (dorsal,  $126 \pm 8 \mu\text{m}$ ,  $n = 5$ ; ventral,  $145 \pm 10 \mu\text{m}$ ,  $n = 6$ ), there was no difference in  $I_h$  current density and  $V_{1/2}$  (Table 3, Fig. 6, dorsal:  $4.7 \pm 0.3 \text{ pA}/\mu\text{m}^2$ ,  $V_{1/2} = -97.9 \pm 5.6 \text{ mV}$ ; ventral:  $4.8 \pm 1.3 \text{ pA}/\mu\text{m}^2$ ,  $V_{1/2} = -101.6 \pm 5.3 \text{ mV}$ ;  $p>0.4$  for both). These results are consistent with the differences found with whole cell recordings.

In adult animals,  $I_h$  and resonance increase in the dendrites with the distance from the soma, and become significantly different from the soma around 200  $\mu\text{m}$  (Magee, 1998; Narayanan and Johnston, 2007; Hu et al., 2009). At PN11-12, dendrites were too thin to be recorded past 150  $\mu\text{m}$ . We thus performed cell attached dendritic recordings a few days later, at PN14-15, a developmental stage when dendrites could be followed in distal stratum radiatum close to the lacunosum-moleculare border (dorsal,  $228 \pm 10 \mu\text{m}$ ,  $n = 9$ ; ventral,  $234 \pm 7 \mu\text{m}$ ,  $n = 7$ ). Cell attached recordings revealed that  $I_h$  density was larger,  $V_{1/2}$  more positive and the activation time constant faster in dorsal than ventral cells (Table 3, Fig. 6, dorsal:  $9.1 \pm 0.8 \text{ pA}/\mu\text{m}^2$ ,  $V_{1/2} = -101.6 \pm 2.6 \text{ mV}$ ,  $\tau = 21.9 \pm 1.9 \text{ ms}$ ; ventral:  $7.2 \pm 0.7 \text{ pA}/\mu\text{m}^2$ ,  $V_{1/2} = -109.0 \pm 3.3 \text{ mV}$ ,  $\tau = 28.4 \pm 2.5 \text{ ms}$ ). For these recordings, we had to use high resistance fine tip electrodes, which prevented us from performing stable whole cell recordings to measure resonance. We conclude that distal dendrites of dorsal cells express a higher density of HCN channels than dendrites of ventral cells.

To determine the molecular correlates of these spatial gradients of  $I_h$  properties, we analyzed the dorso-ventral pattern of expression of HCN1 and HCN2 subunits, the two major contributors to hippocampal  $I_h$  (Ludwig et al., 2003; Nolan et al., 2004; Surges et al., 2006; Brewster et al., 2007) at mRNA and protein levels, and at several levels of resolution.

### Increased levels of HCN1, HCN2 and TRIP8b in dorsal hippocampus

Using RT-PCR, we did not find differences in HCN1 and HCN2 mRNA levels between ventral and dorsal hippocampus (Fig. 7A,  $n=4$ ). Western blot analyses revealed increased levels of HCN1 and HCN2 protein in dorsal as compared to ventral hippocampus (Fig. 7B, expressed as a fraction of ventral hippocampal levels, HCN1: dorsal  $1.38 \pm 0.04$ , ventral  $1.00 \pm 0.16$ ,  $p<0.04$ ; HCN2: dorsal  $2.39 \pm 0.22$ , ventral  $1.00 \pm 0.18$ ,  $p<0.0006$ ). Immunocytochemistry, enabling visualization of the cellular origin and subcellular distribution of HCN1, revealed a clear dorso-ventral gradient of this channel (Fig. 8). When the optical density (OD) of the HCN1 signal was quantified per that of the dendritic marker MAP2 (Fig. 8), increased density of HCN1/dendrite was found in dorsal vs. ventral hippocampus. Remarkably, this dorsal augmentation was confined to the distal dendritic

fields in stratum lacunosum moleculare (Fig. 8), in keeping with the electrophysiology data (Fig. 6).

### Increased TRIP8b levels in the ventral hippocampus

Increased HCN protein levels are consistent with increased  $I_h$  amplitude in dorsal cell somata and dendrites. However, additional factors, including interactions with auxiliary proteins such as TRIP8b, influence  $I_h$  properties (Santoro et al., 2004; Lewis et al., 2009; Zolles et al., 2009). TRIP8b was increased five-fold in the dorsal as compared to the ventral hippocampus (Fig. 7B, dorsal  $5.00 \pm 0.87$ , ventral  $1.00 \pm 0.32$ ,  $p < 0.002$ ). Because TRIP8b inhibits cAMP modulation of  $I_h$  (Zolles et al., 2009), we predicted that HCN-dependent properties would be more sensitive to cAMP modulation in ventral than dorsal cells, where TRIP8b levels are higher.

### $I_h$ and resonance show increased sensitivity to cAMP in the ventral hippocampus

Intracellular application of 10  $\mu$ M cAMP increased resonance in ventral cells (Fig. 9A;  $n=8$ ,  $F_{res}$   $3.7 \pm 0.2$  Hz before cAMP,  $4.3 \pm 0.2$  Hz after cAMP;  $Q$   $1.13 \pm 0.01$  before,  $1.19 \pm 0.01$  after);  $I_h$  amplitude (Fig. 9B; at  $-140$  mV,  $256 \pm 23$  pA before,  $379 \pm 42$  pA after; at  $-90$  mV,  $119 \pm 16$  pA before,  $188 \pm 17$  pA after), shifted  $V_{1/2}$  by 9 mV toward more depolarized values (Fig. 9B;  $-91.2 \pm 1.2$  mV before,  $-82.5 \pm 1.9$  mV after) and decreased the fast activation time constant (Fig. 9B;  $\tau_{fast}$ ,  $28 \pm 1$  ms before,  $23 \pm 2$  ms after;  $\tau_{slow}$ ,  $225 \pm 28$  ms before,  $215 \pm 27$  ms after). In dorsal cells (Fig. 9;  $n=15$ ), apart from a significant shift of  $V_{1/2}$ , there were no statistically significant modifications in the other parameters ( $I_h$  amplitude at  $-140$  mV,  $313 \pm 25$  pA before,  $304 \pm 30$  pA after; at  $-90$  mV,  $150 \pm 11$  pA before,  $159 \pm 16$  pA after;  $V_{1/2}$   $-88.9 \pm 0.7$  mV before,  $-82.5 \pm 1.7$  mV after;  $\tau_{fast}$ ,  $17 \pm 2$  ms before,  $19 \pm 1$  ms after;  $\tau_{slow}$ ,  $195 \pm 27$  ms before,  $168 \pm 22$  ms after;  $F_{res}$   $4.4 \pm 0.2$  Hz before,  $4.3 \pm 0.2$  Hz after;  $Q$   $1.27 \pm 0.02$  before,  $1.25 \pm 0.03$  after; paired t-test  $p > 0.4$  for all parameters, except for  $V_{1/2}$  for which  $p < 0.005$ ).

Finally, cAMP decreased temporal summation of EPSPs in ventral cells (Fig. 9C, at 25 Hz, before cAMP  $7 \pm 3\%$ , after cAMP  $0 \pm 1\%$ ; at 50 Hz, before  $72 \pm 5\%$ , after  $60 \pm 4\%$ ,  $n=12$ ) but not in dorsal cells (Fig. 9D, at 25 Hz, before  $-3 \pm 1\%$ , after  $-4 \pm 2\%$ ; at 50 Hz, before  $41 \pm 4\%$ , after  $45 \pm 4\%$ ,  $n=14$ ). Addition of ZD7288 at the end of the experiment equalized temporal summation in dorsal and ventral cells (at 25 Hz, dorsal  $58 \pm 8\%$ ,  $n=6$ ; ventral  $57 \pm 23\%$ ,  $n=4$ ; at 50 Hz, dorsal  $140 \pm 26\%$ ,  $n=6$ ; ventral  $151 \pm 25\%$ ,  $n=4$ , not shown).

## Discussion

Our results reveal a dorso-ventral gradient during the second postnatal week, characterized by augmented resonance and  $I_h$  and lower temporal summation in dorsal than ventral cells. This indicates an early and fine-tuned orchestration of hippocampal organization. Regulated molecular programs of HCN1/2 and TRIP8b expression seem to contribute to this organization.

The three-dimensional organization of the medial entorhinal cortex allows the functional mapping of stellate cells along the dorso-ventral axis. The curvature of the hippocampus prevents such an approach, since dendritic recordings require a complete preservation of the cell, which is obtained in one or two slices per hemisphere, hence the use of dorsal and ventral slices in this study. The hippocampus develops in a dorso-ventral gradient (Bayer, 1980). Dorsal cells could thus be more mature than ventral ones at PN11-12. However, the morphological characteristics of the recorded cells were similar in the dorsal and ventral hippocampus.

Results obtained in young (present study) and adult animals (Narayanan and Johnston, 2007; Marcelin et al., 2009) show that resonance increases with age in CA1 pyramidal cells, in parallel with the frequency of theta rhythm recorded *in vivo* (Leblanc and Bland, 1979). At PN5-6, there is no resonance and no proper theta oscillations *in vivo*. At PN11,  $F_{res} \approx 4$  Hz, and theta frequency is 3.4–3.9 Hz *in vivo*; whereas in adult animals,  $F_{res} = 6$ –10 Hz along the somato-dendritic axis, and theta frequency is 4–12 Hz. This does not necessarily imply that a causal relationship exists between resonance frequency and theta rhythm. We suggest that resonance parameters at the single cell level are tuned to the frequency generated at the network level. Interestingly, phase lead, which may be important for phase precession (Narayanan and Johnston, 2008; Marcelin et al., 2009), was below the theta frequency range at PN11-12, but in the theta range in adult animals (Narayanan and Johnston, 2008; Marcelin et al., 2009). This may be related to the fact that animals poorly explore actively their environment at PN11-12. In adult animals, resonance is much stronger in both hippocampus (Narayanan and Johnston, 2007; Marcelin et al., 2009) and entorhinal cortex (Giocomo et al., 2007; Garden et al., 2008; Giocomo and Hasselmo, 2008; Boehlen et al., 2010).

Resonance and phase responses were larger, and temporal summation smaller, in dorsal than ventral somata, suggesting that CA1 principal cells process information differently along the dorso-ventral axis. Whether such topological arrangement plays a role in the dorso-ventral expansion of place fields remains to be determined. The differences in  $I_h$  properties are sufficient to account for the differences in resonance and temporal summation between dorsal and ventral cells, since a decrease in  $I_h$  amplitude and/or availability decreases resonance and increases temporal summation (Narayanan and Johnston, 2007; Marcelin et al., 2009). The developmental profile of  $I_h$  properties is similar to the developmental profile of resonance, as at early stages  $I_h$  has slow kinetics and small amplitude (Surges et al., 2006), leading to a lack of resonance. Interestingly, dentate granule cells do not appear to express  $I_h$  at early stages (Stabel et al., 1992). The spatial profile of resonance, temporal summation and  $I_h$  in CA1 pyramidal cells is thus similar to that found in stellate cells of the entorhinal cortex (Giocomo et al., 2007; Garden et al., 2008; Giocomo and Hasselmo, 2008; Boehlen et al., 2010). In our conditions ( $-70$  mV), resonance and temporal coding were  $I_h$ -dependent. Near firing threshold, resonance involves other sets of channels, in particular  $I_M$  and  $I_{NaP}$  (Hutcheon and Yarom, 2000; Hu et al., 2009). The dorso-ventral distributions of  $I_M$  and  $I_{NaP}$  remain to be assessed.

At PN11-12,  $I_h$  and resonance properties were not larger in dendrites than in somata. Since dendritic recordings could not be performed farther than  $125 \mu\text{m}$  away from the soma and since resonance and  $I_h$  starts to increase farther than  $150 \mu\text{m}$  in adult animals (Magee, 1998; Narayanan and Johnston, 2007), we may have not recorded far enough to see a difference. However, dendritic recordings performed far enough from the soma at PN14-15 revealed a clear difference between the dorsal and ventral part of the hippocampus.

Our attempt to find molecular correlates revealed an increased expression of HCN1 and TRIP8b in dorsal hippocampus, which could account for larger  $I_h$  current and decreased cAMP sensitivity in dorsal cells (Santoro et al., 2004; Lewis et al., 2009; Zolles et al., 2009). Interestingly, cAMP shifted  $V_{1/2}$  but not the amplitude of  $I_h$  in dorsal cells. The shift of  $V_{1/2}$  by cAMP was not sufficient to change resonance, as predicted by modeling (Narayanan and Johnston, 2007). In contrast, cAMP increased resonance and decreased temporal summation in ventral somata. Such modulation could play an important role when cAMP levels are specifically increased in the ventral hippocampus during strong emotional conditions (Klamer et al., 2005). Relevant to this concept of functional segregation, resonance and  $I_h$  are differentially modulated by cholinergic inputs in stellate cells along the dorso-ventral axis (Heys et al., 2010). Increased levels of HCN2 and TRIP8b in the dorsal hippocampus

should have resulted in slower  $I_h$  kinetics in dorsal cells, in contrast to the present findings. Other putative modulators of  $I_h$ , including MiRP1 (Proenza et al., 2002), phosphoinositides (Zolles et al., 2006; Pian et al., 2007), and protein kinases (Poolos et al., 2006), might be involved. Their dorso-ventral expression patterns remain to be assessed. The larger expression of HCN2 in the dorsal hippocampus should have resulted in a larger modulation by cAMP. The high expression of *Trip8b* may play a role in this decreased sensitivity, a proposal that can be tested in *Trip8b* invalidated animals.

To conclude, our data support the concept that physiological architecture is tuned to the information to be processed (Giocomo et al., 2007; Garden et al., 2008). We propose the existence of organizational rules linking resonance,  $I_h$ , and perhaps theta oscillations (although causality between these phenomena remains to be established). Given the similarity of the spatial and temporal profiles of these parameters in stellate cells and CA1 pyramidal cells, we propose that the developmental programs controlling these parameters may be shared by the entorhinal cortex and the hippocampus, two structures processing complementary features of the environment.

## Acknowledgments

This work was supported by INSERM, ANR ANTARES and MINOS grants, EPICURE (C.B. and A.B.), NIH 1F30NS064757 (A.S.L.) and 1K02NS05595 1R01NS059934 (D.M.C.), T32 04450 (S.M) and R37 NS35439 (T.Z.B). We thank M.-P. Nesa and A. Ghestem for technical help. B.M. was supported by CIFRE contract with Elekta.

## Reference List

- Bayer SA. Development of the hippocampal region in the rat. II. Morphogenesis during embryonic and early postnatal life. *J Comp Neurol.* 1980; 190:115–134. [PubMed: 7381049]
- Bender RA, Brewster A, Santoro B, Ludwig A, Hofmann F, Biel M, Baram TZ. Differential and age-dependent expression of hyperpolarization-activated, cyclic nucleotide-gated cation channel isoforms 1–4 suggests evolving roles in the developing rat hippocampus. *Neuroscience.* 2001; 106:689–698. [PubMed: 11682156]
- Bender RA, Galindo R, Mameli M, Gonzalez-Vega R, Valenzuela CF, Baram TZ. Synchronized network activity in developing rat hippocampus involves regional hyperpolarization-activated cyclic nucleotide-gated (HCN) channel function. *Eur J Neurosci.* 2005; 22:2669–2674. [PubMed: 16307610]
- Bender RA, Kirschstein T, Kretz O, Brewster AL, Richichi C, Ruschenschmidt C, Shigemoto R, Beck H, Frotscher M, Baram TZ. Localization of HCN1 channels to presynaptic compartments: novel plasticity that may contribute to hippocampal maturation. *J Neurosci.* 2007; 27:4697–4706. [PubMed: 17460082]
- Bernard C, Anderson A, Becker A, Poolos NP, Beck H, Johnston D. Acquired dendritic channelopathy in temporal lobe epilepsy. *Science.* 2004; 305:532–535. [PubMed: 15273397]
- Boehlen A, Heinemann U, Erchova I. The range of intrinsic frequencies represented by medial entorhinal cortex stellate cells extends with age. *J Neurosci.* 2010; 30:4585–4589. [PubMed: 20357109]
- Brewster AL, Chen Y, Bender RA, Yeh A, Shigemoto R, Baram TZ. Quantitative Analysis and Subcellular Distribution of mRNA and Protein Expression of the Hyperpolarization-Activated Cyclic Nucleotide-Gated Channels throughout Development in Rat Hippocampus. *Cereb Cortex.* 2007; 17:702–712. [PubMed: 16648453]
- Burton BG, Economo MN, Lee GJ, White JA. Development of theta rhythmicity in entorhinal stellate cells of the juvenile rat. *J Neurophysiol.* 2008; 100:3144–3157. [PubMed: 18829850]
- Chen Y, Bender RA, Frotscher M, Baram TZ. Novel and transient populations of corticotropin-releasing hormone-expressing neurons in developing hippocampus suggest unique functional roles: a quantitative spatiotemporal analysis. *J Neurosci.* 2001; 21:7171–7181. [PubMed: 11549728]

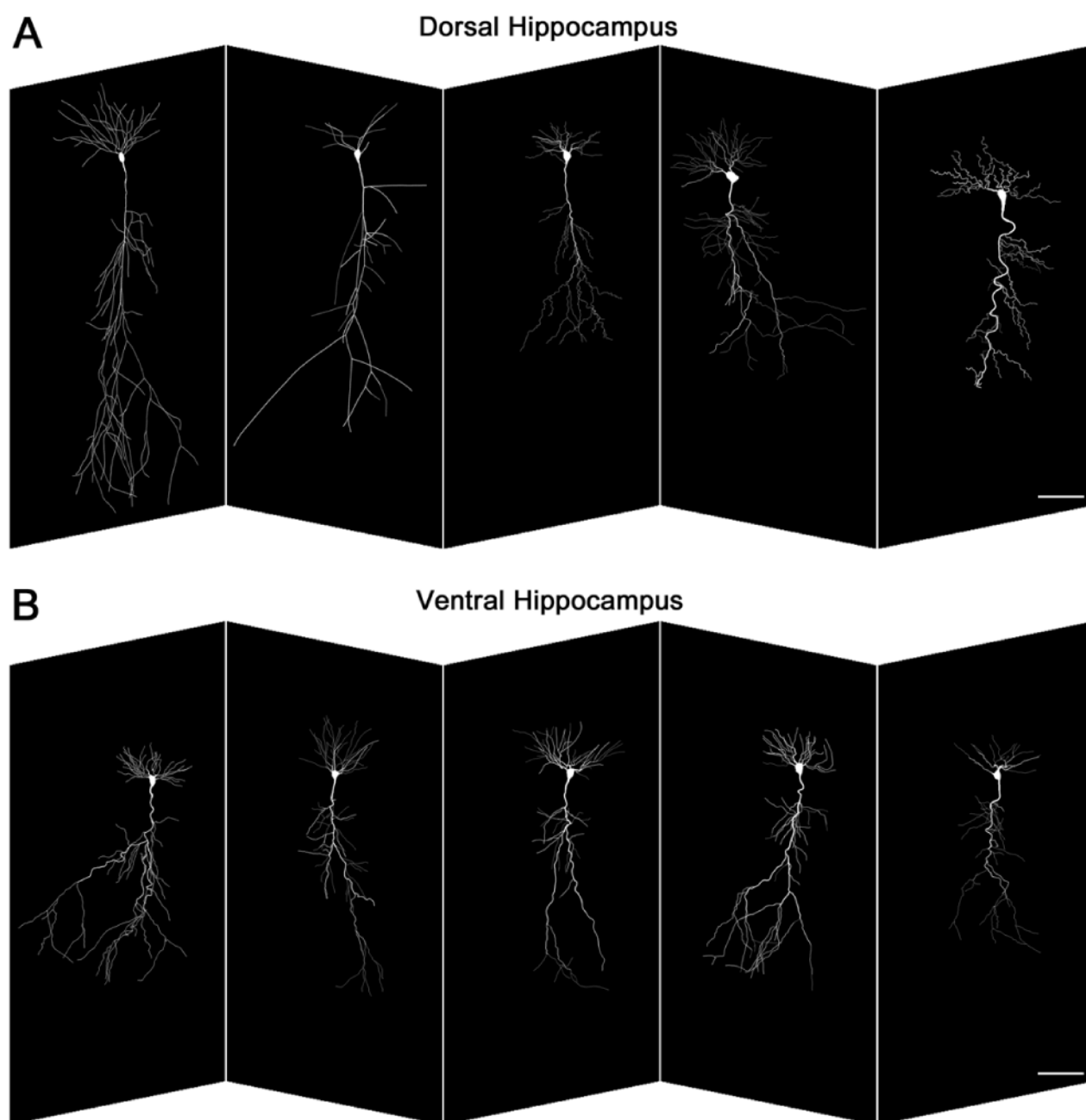
- Dong HW, Swanson LW, Chen L, Fanselow MS, Toga AW. Genomic-anatomic evidence for distinct functional domains in hippocampal field CA1. *Proc Natl Acad Sci U S A*. 2009; 106:11794–11799. [PubMed: 19561297]
- Esclapez M, Hirsch JC, Ben-Ari Y, Bernard C. Newly formed excitatory pathways provide a substrate for hyperexcitability in experimental temporal lobe epilepsy. *J Comp Neurol*. 1999; 408:449–460. [PubMed: 10340497]
- Fanselow MS, Dong HW. Are the dorsal and ventral hippocampus functionally distinct structures? *Neuron*. 2010; 65:7–19. [PubMed: 20152109]
- Fink L, Seeger W, Ermert L, Hanze J, Stahl U, Grimminger F, Kummer W, Bohle RM. Real-time quantitative RT-PCR after laser-assisted cell picking. *Nat Med*. 1998; 4:1329–1333. [PubMed: 9809560]
- Garden DL, Dodson PD, O'Donnell C, White MD, Nolan MF. Tuning of synaptic integration in the medial entorhinal cortex to the organization of grid cell firing fields. *Neuron*. 2008; 60:875–889. [PubMed: 19081381]
- Giocomo LM, Hasselmo ME. Time constants of h current in layer ii stellate cells differ along the dorsal to ventral axis of medial entorhinal cortex. *J Neurosci*. 2008; 28:9414–9425. [PubMed: 18799674]
- Giocomo LM, Zilli EA, Fransen E, Hasselmo ME. Temporal frequency of subthreshold oscillations scales with entorhinal grid cell field spacing. *Science*. 2007; 315:1719–1722. [PubMed: 17379810]
- Heys JG, Giocomo LM, Hasselmo ME. Cholinergic modulation of the resonance properties of stellate cells in layer II of medial entorhinal cortex. *J Neurophysiol*. 2010; 104:258–270. [PubMed: 20445030]
- Hines ML, Carnevale NT. The NEURON simulation environment. *Neural Comput*. 1997; 9:1179–1209. [PubMed: 9248061]
- Hu H, Vervaeke K, Graham LJ, Storm JF. Complementary theta resonance filtering by two spatially segregated mechanisms in CA1 hippocampal pyramidal neurons. *J Neurosci*. 2009; 29:14472–14483. [PubMed: 19923281]
- Hu H, Vervaeke K, Storm JF. Two forms of electrical resonance at theta frequencies, generated by M-current, h-current and persistent Na<sup>+</sup> current in rat hippocampal pyramidal cells. *J Physiol*. 2002; 545:783–805. [PubMed: 12482886]
- Hutcheon B, Yarom Y. Resonance, oscillation and the intrinsic frequency preferences of neurons. *Trends Neurosci*. 2000; 23:216–222. [PubMed: 10782127]
- Jung S, Warner LN, Pitsch J, Becker AJ, Poolos NP. Rapid loss of dendritic HCN channel expression in hippocampal pyramidal neurons following status epilepticus. *J Neurosci*. 2011; 31:14291–14295. [PubMed: 21976514]
- Kjelstrup KB, Solstad T, Brun VH, Hafting T, Leutgeb S, Witter MP, Moser EI, Moser MB. Finite scale of spatial representation in the hippocampus. *Science*. 2008; 321:140–143. [PubMed: 18599792]
- Klamer D, Palsson E, Fejgin K, Zhang J, Engel JA, Svensson L. Activation of a nitric-oxide-sensitive cAMP pathway with phencyclidine: elevated hippocampal cAMP levels are temporally associated with deficits in prepulse inhibition. *Psychopharmacology (Berl)*. 2005; 179:479–488. [PubMed: 15619121]
- Langston RF, Ainge JA, Couey JJ, Canto CB, Bjerknes TL, Witter MP, Moser EI, Moser MB. Development of the spatial representation system in the rat. *Science*. 2010; 328:1576–1580. [PubMed: 20558721]
- Leblanc MO, Bland BH. Developmental aspects of hippocampal electrical activity and motor behavior in the rat. *Exp Neurol*. 1979; 66:220–237. [PubMed: 226389]
- Lewis AS, Schwartz E, Chan CS, Noam Y, Shin M, Wadman WJ, Surmeier DJ, Baram TZ, Macdonald RL, Chetkovich DM. Alternatively spliced isoforms of TRIP8b differentially control h channel trafficking and function. *J Neurosci*. 2009; 29:6250–6265. [PubMed: 19439603]
- Ludwig A, Budde T, Stieber J, Moosmang S, Wahl C, Holthoff K, Langebartels A, Wotjak C, Munsch T, Zong X, Feil S, Feil R, Lancel M, Chien KR, Konnerth A, Pape HC, Biel M, Hofmann F. Absence epilepsy and sinus dysrhythmia in mice lacking the pacemaker channel HCN2. *EMBO J*. 2003; 22:216–224. [PubMed: 12514127]



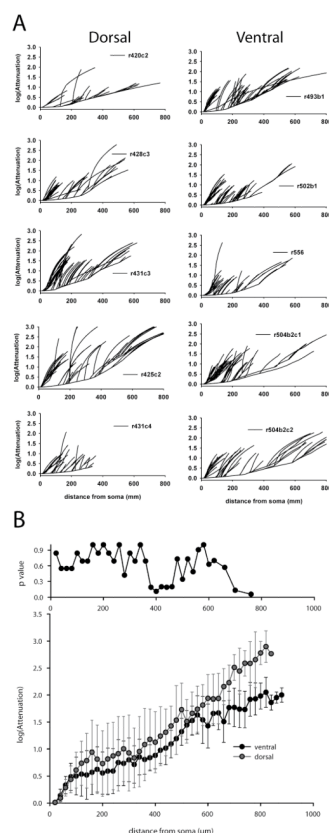
- Magee JC. Dendritic hyperpolarization-activated currents modify the integrative properties of hippocampal CA1 pyramidal neurons. *J Neurosci*. 1998; 18:7613–7624. [PubMed: 9742133]
- Marcelin B, Chauviere L, Becker A, Migliore M, Esclapez M, Bernard C. h channel-dependent deficit of theta oscillation resonance and phase shift in temporal lobe epilepsy. *Neurobiol Dis*. 2009; 33:436–447. [PubMed: 19135151]
- Moser MB, Moser EI. Functional differentiation in the hippocampus. *Hippocampus*. 1998; 8:608–619. [PubMed: 9882018]
- Narayanan R, Johnston D. Long-term potentiation in rat hippocampal neurons is accompanied by spatially widespread changes in intrinsic oscillatory dynamics and excitability. *Neuron*. 2007; 56:1061–1075. [PubMed: 18093527]
- Narayanan R, Johnston D. The h channel mediates location dependence and plasticity of intrinsic phase response in rat hippocampal neurons. *J Neurosci*. 2008; 28:5846–5850. [PubMed: 18509046]
- Nolan MF, Dudman JT, Dodson PD, Santoro B. HCN1 channels control resting and active integrative properties of stellate cells from layer II of the entorhinal cortex. *J Neurosci*. 2007; 27:12440–12451. [PubMed: 18003822]
- Nolan MF, Malleret G, Dudman JT, Buhl DL, Santoro B, Gibbs E, Vronskaya S, Buzsaki G, Siegelbaum SA, Kandel ER, Morozov A. A behavioral role for dendritic integration: HCN1 channels constrain spatial memory and plasticity at inputs to distal dendrites of CA1 pyramidal neurons. *Cell*. 2004; 119:719–732. [PubMed: 15550252]
- O'Keefe, J.; Nadel, L. *The hippocampus as a cognitive map*. Oxford: Oxford University; 1978.
- Pian P, Bucchi A, Decostanzo A, Robinson RB, Siegelbaum SA. Modulation of cyclic nucleotide-regulated HCN channels by PIP(2) and receptors coupled to phospholipase C. *Pflugers Arch*. 2007; 455:125–145. [PubMed: 17605039]
- Pike FG, Goddard RS, Suckling JM, Ganter P, Kasthuri N, Paulsen O. Distinct frequency preferences of different types of rat hippocampal neurones in response to oscillatory input currents. *J Physiol*. 2000; 529(Pt 1):205–213. [PubMed: 11080262]
- Poolos NP, Bullis JB, Roth MK. Modulation of h-channels in hippocampal pyramidal neurons by p38 mitogen-activated protein kinase. *J Neurosci*. 2006; 26:7995–8003. [PubMed: 16870744]
- Proenza C, Angoli D, Agranovich E, Macri V, Accili EA. Pacemaker channels produce an instantaneous current. *J Biol Chem*. 2002; 277:5101–5109. [PubMed: 11741901]
- Santoro B, Wainger BJ, Siegelbaum SA. Regulation of HCN channel surface expression by a novel C-terminal protein-protein interaction. *J Neurosci*. 2004; 24:10750–10762. [PubMed: 15564593]
- Sargolini F, Fyhn M, Hafting T, McNaughton BL, Witter MP, Moser MB, Moser EI. Conjunctive representation of position, direction, and velocity in entorhinal cortex. *Science*. 2006; 312:758–762. [PubMed: 16675704]
- Shin M, Chetkovich DM. Activity-dependent regulation of h channel distribution in hippocampal CA1 pyramidal neurons. *J Biol Chem*. 2007; 282:33168–33180. [PubMed: 17848552]
- Shin M, Simkin D, Suyeoka GM, Chetkovich DM. Evaluation of HCN2 abnormalities as a cause of juvenile audiogenic seizures in Black Swiss mice. *Brain Res*. 2006; 1083:14–20. [PubMed: 16542642]
- Stabel J, Ficker E, Heinemann U. Young CA1 pyramidal cells of rats, but not dentate gyrus granule cells, express a delayed inward rectifying current with properties of IQ. *Neurosci Lett*. 1992; 135:231–234. [PubMed: 1378215]
- Surges R, Brewster AL, Bender RA, Beck H, Feuerstein TJ, Baram TZ. Regulated expression of HCN channels and cAMP levels shape the properties of the h current in developing rat hippocampus. *Eur J Neurosci*. 2006; 24:94–104. [PubMed: 16882011]
- Swanson L, Wyss J, Cowan W. An autoradiographic study of the organization of intrahippocampal association pathways in the rat. *J Comp Neurol*. 1978; 181:681–715. [PubMed: 690280]
- Thompson CL, Pathak SD, Jeromin A, Ng LL, MacPherson CR, Mortrud MT, Cusick A, Riley ZL, Sunkin SM, Bernard A, Puchalski RB, Gage FH, Jones AR, Bajic VB, Hawrylycz MJ, Lein ES. Genomic anatomy of the hippocampus. *Neuron*. 2008; 60:1010–1021. [PubMed: 19109908]



- Vasilyev DV, Barish ME. Postnatal development of the hyperpolarization-activated excitatory current  $I_h$  in mouse hippocampal pyramidal neurons. *J Neurosci.* 2002; 22:8992–9004. [PubMed: 12388606]
- Wills TJ, Cacucci F, Burgess N, O'Keefe J. Development of the hippocampal cognitive map in preweanling rats. *Science.* 2010; 328:1573–1576. [PubMed: 20558720]
- Yoshida M, Giocomo LM, Boardman I, Hasselmo ME. Frequency of subthreshold oscillations at different membrane potential voltages in neurons at different anatomical positions on the dorsoventral axis in the rat medial entorhinal cortex. *J Neurosci.* 2011; 31:12683–12694. [PubMed: 21880929]
- Zolles G, Klocker N, Wenzel D, Weisser-Thomas J, Fleischmann BK, Roeper J, Fakler B. Pacemaking by HCN channels requires interaction with phosphoinositides. *Neuron.* 2006; 52:1027–1036. [PubMed: 17178405]
- Zolles G, Wenzel D, Bildl W, Schulte U, Hofmann A, Muller CS, Thumfart JO, Vlachos A, Deller T, Pfeifer A, Fleischmann BK, Roeper J, Fakler B, Klocker N. Association with the auxiliary subunit PEX5R/Trip8b controls responsiveness of HCN channels to cAMP and adrenergic stimulation. *Neuron.* 2009; 62:814–825. [PubMed: 19555650]

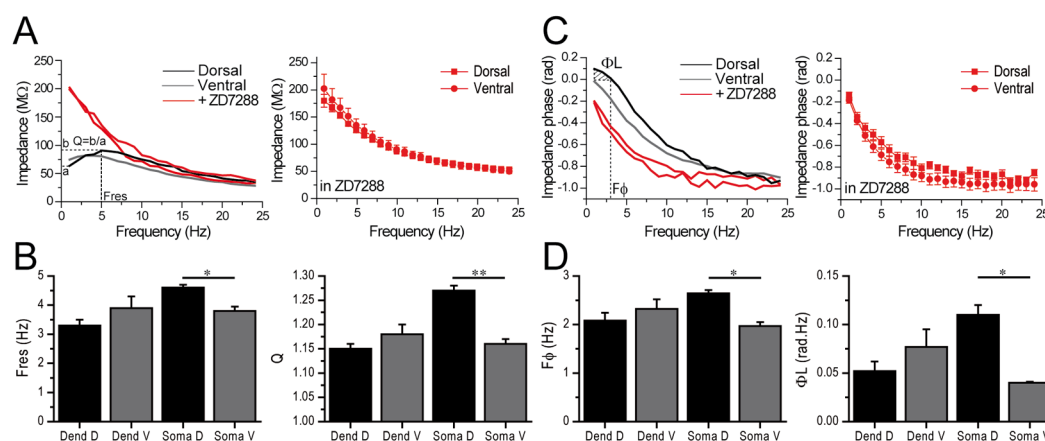


**Figure 1.** Tridimensional reconstruction of CA1 pyramidal cells recorded in the dorsal (n=5) and ventral (n=5) parts of the hippocampus. There are no overt morphological differences (see Table 1 for quantification).



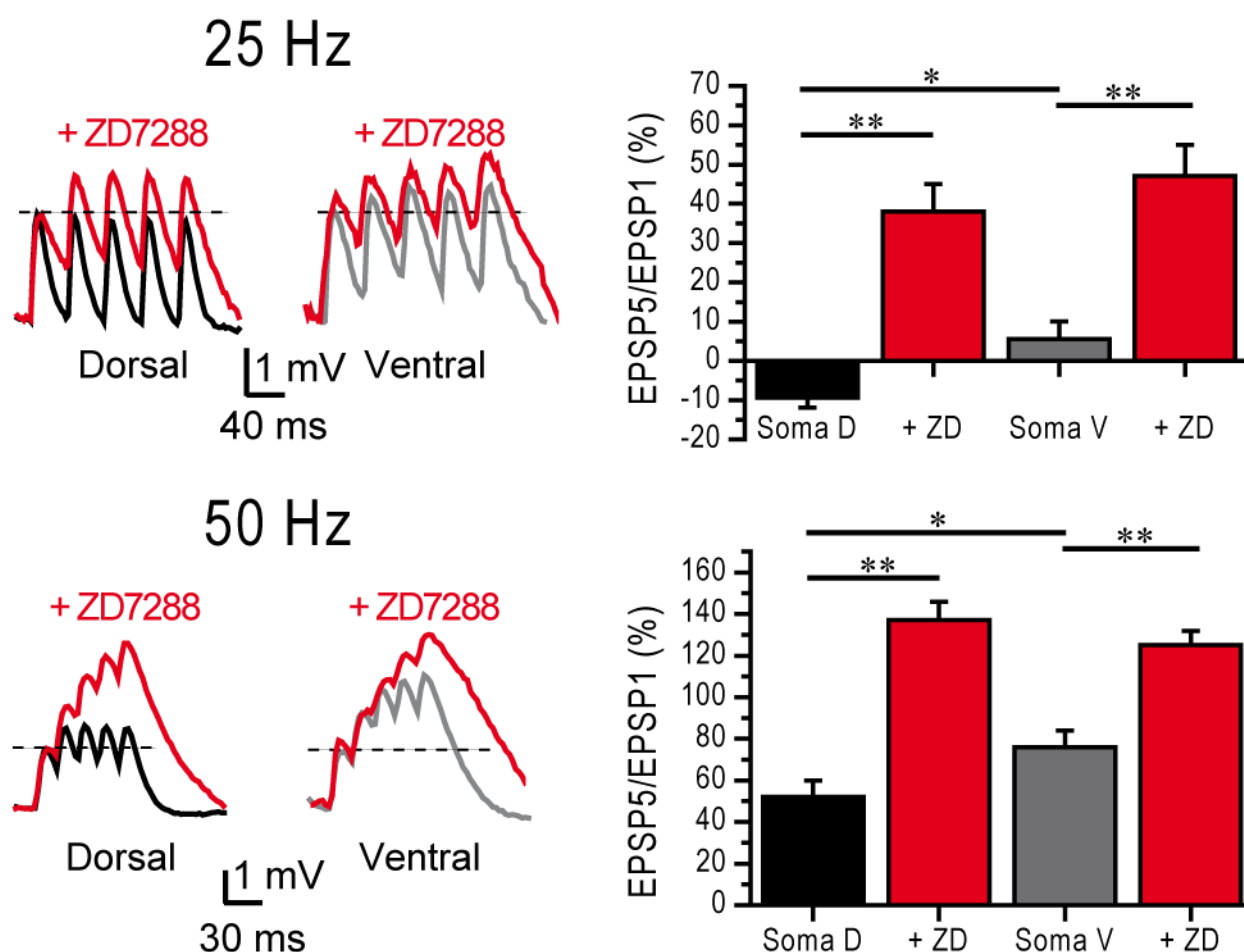
**Figure 2.**

**A**, Curves for each neuron shown in Figure 1. Voltage attenuation appears homogeneous between the different CA1 pyramidal cells. **B**, Average voltage attenuation (log scale,  $\pm$  s.d.) for the reconstructed ventral and dorsal CA1 pyramidal cells. There was no statistical difference between the average values of attenuation, calculated at different distances from the soma, from cells recorded in the ventral and dorsal parts of the hippocampus (Mann-Whitney Rank Sum Test for  $n > 3$ , p value as a function of distance is plotted in the top panel).



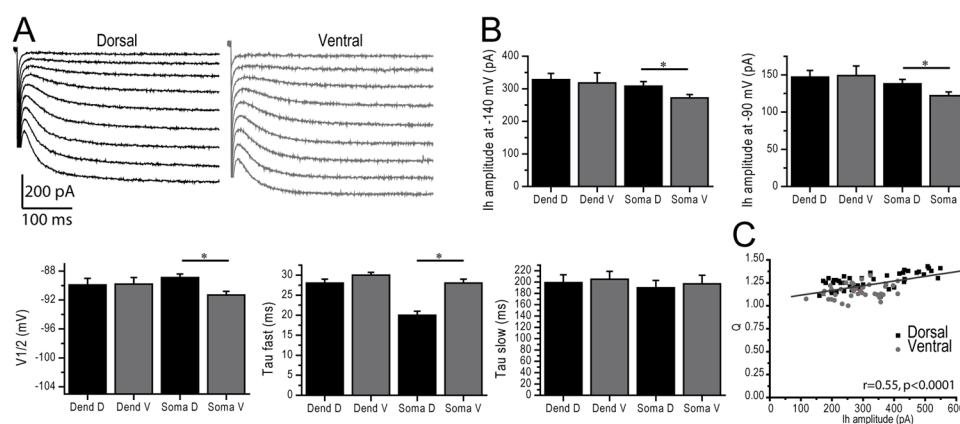
**Figure 3.**

Resonance is stronger in ventral than in dorsal CA1 pyramidal cell somata. **A**, Left panel shows the impedance magnitude of a dorsal (black curve) and ventral (grey curve) soma before and after application of ZD7288 (red curves). Note that resonance is smaller in the ventral as compared to the dorsal cell. Resonance was abolished by ZD7288. Right panel, average impedance curves are identical in dorsal and ventral somata after ZD7288 (ANOVA for group and frequency as factors). **B**, Histograms of resonance parameters. The resonance frequency ( $F_{res}$ ) and the amplification ratio ( $Q$ ) were similar in ventral (V) and dorsal (D) dendrites. In somata,  $F_{res}$  and  $Q$  were larger in dorsal than in ventral cells. **C**, Left panel, phase responses in the same cells as in (A). Right panel, average phase responses are similar in dorsal and ventral cells after ZD7288 (ANOVA for group and frequency as factors). **D**, Histograms of phase responses.  $F_{\phi}$  and  $\Phi_L$  are larger in dorsal than in ventral somata, but not in dendrites. \*,  $p < 0.05$ ; \*\*,  $p < 0.01$ .



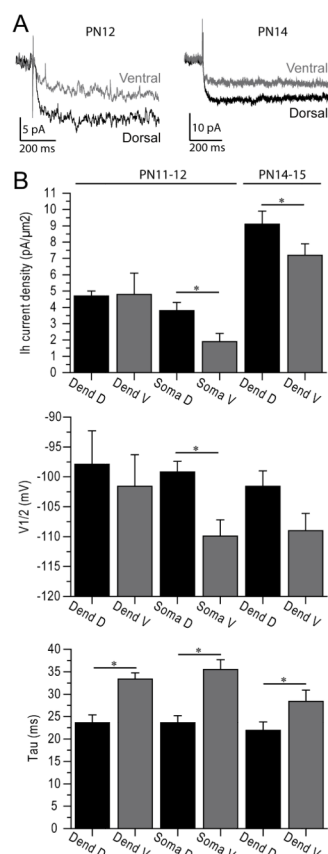
**Figure 4.**

Temporal summation is larger in ventral than is dorsal cells. Left panels, examples of temporal summation at 25 Hz and 50 Hz in a dorsal (black) and ventral (grey) soma. Summation was increased in the presence of ZD7288. Right panels, histograms of summation ratios for 25 Hz and 50 Hz trains in dorsal and ventral cells. Blockade of  $I_h$  normalized summation in dorsal and ventral cells. ANOVA \*,  $p < 0.05$ ; \*\*,  $p < 0.001$ .

**Figure 5.**

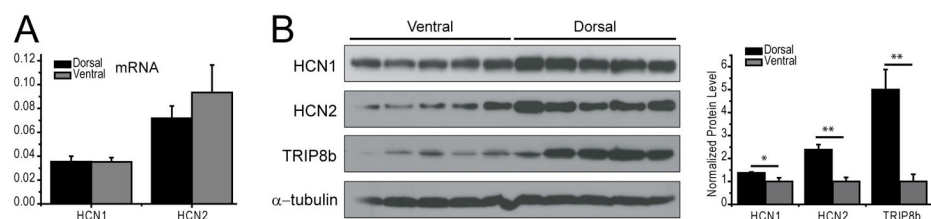
$I_h$  availability is increased in dorsal as compared to ventral somata. **A**, Left (dorsal soma) and right (ventral soma) panel:  $I_h$  activation (whole cell recordings). Note the increased amplitude in the dorsal cell. **B**, Summary histograms of  $I_h$  amplitude at  $-140$  and  $-90$  mV (whole cell recordings),  $V_{1/2}$  and activation time constant. The amplitude of  $I_h$  current is larger, and kinetics faster in dorsal than ventral somata. In contrast,  $I_h$  properties measured in proximal dendrites are similar. **C**, Relationship between  $I_h$  amplitude and the amplification ratio  $Q$  in dorsal (filled squares) and ventral (filled circles) somata. The whole dataset showed a linear relationship between  $I_h$  and  $Q$ . \*,  $p<0.05$





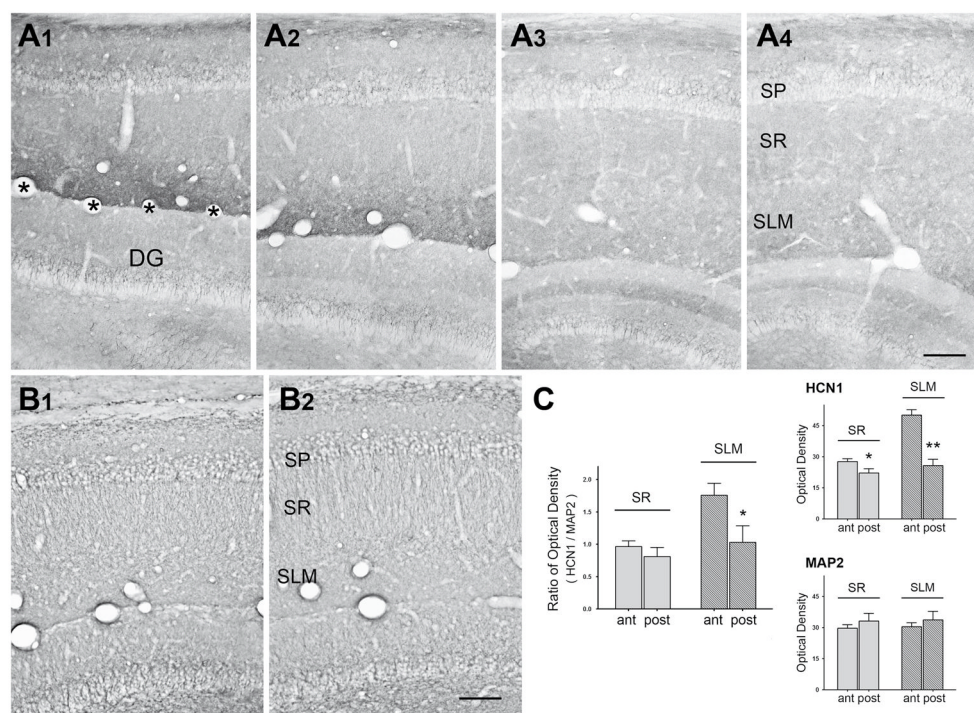
**Figure 6.**

$I_h$  properties measured with cell attached recordings. **A**, Left panel, examples of  $I_h$  activation using cell attached recordings of a dorsal and ventral soma at PN12. Right panel, examples of  $I_h$  activation using cell attached recordings of a dorsal (250  $\mu m$  from the soma) and ventral (260  $\mu m$  from the soma) dendrite at PN14. Channel currents are larger in the dorsal part of the hippocampus. **B**, Summary histograms of  $I_h$  current density,  $V_{1/2}$  and tau in somata and dendrites recorded at PN11-12, and in dendrites at PN14-15. Somatic recordings revealed larger  $I_h$  with faster kinetics in dorsal than ventral somata at PN11-12. In contrast, dendritic values were not different apart from the activation time constant. The lack of difference may due to the fact that only proximal recordings (around 130  $\mu m$ ) could be performed at that stage of development. The difference in the dendrites became significant at PN14-15, as recordings could be performed around 230  $\mu m$ . \*,  $p < 0.05$ .



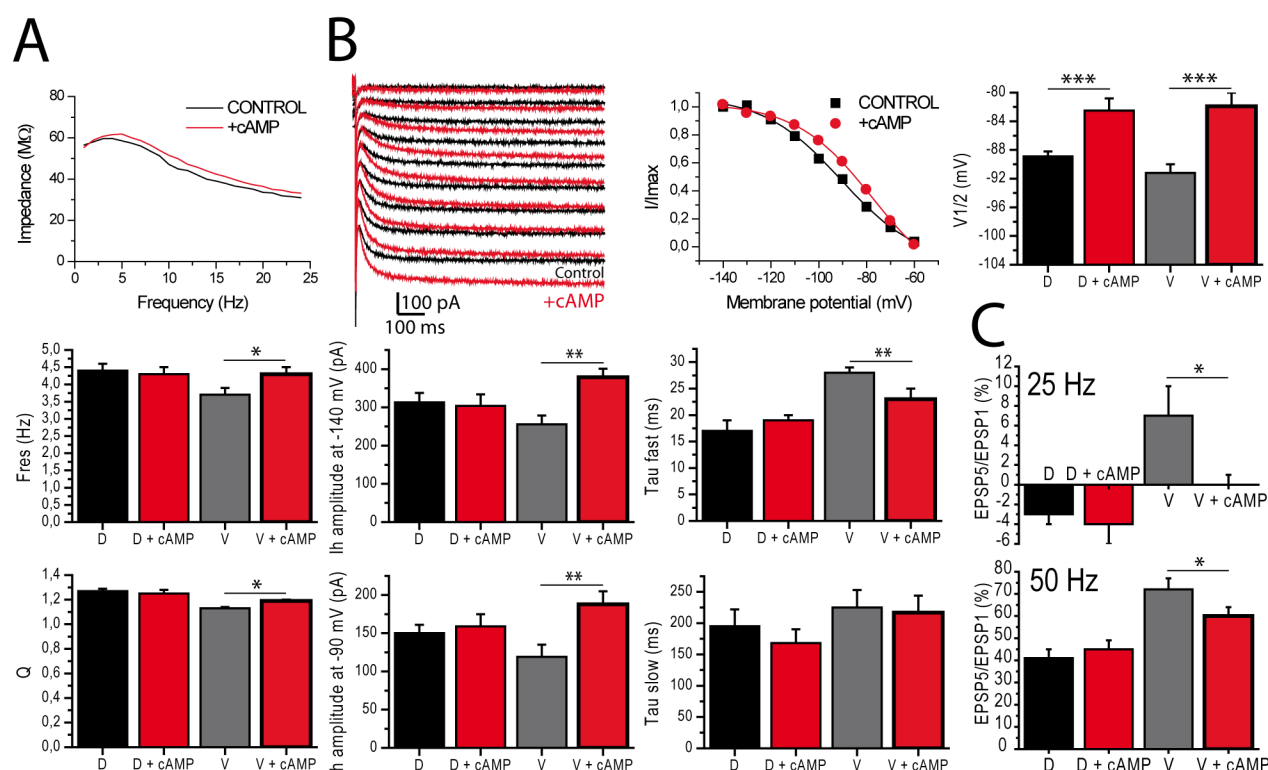
**Figure 7.**

HCN subunit and TRIP8b dorso-ventral differential distribution as a molecular substrate for the larger  $I_h$  amplitude in dorsal versus ventral somata. Right panel: **A**, RT-PCR did not reveal dorso-ventral differences for HCN1 and HCN2 mRNA. **B**, HCN1, HCN2, and TRIP8b protein levels from western blots demonstrate that HCN1, HCN2 and TRIP8b protein levels are significantly larger in dorsal than ventral hippocampus. \*,  $p < 0.05$ ; \*\*,  $p < 0.001$ .



**Figure 8.**

Dorsal-ventral gradient of dendritic HCN1 channels along developing hippocampal CA1. **A**, In the developing hippocampus (P12), HCN1 channels are localized in the dendritic field of CA1 pyramidal cells including strata radiatum (SR) and lacunosum-moleculare (SLM). The HCN1-immunoreactivity is higher in the dorsal level (A1, Bregma:  $\sim -3.0$ mm), then decreases gradually along the dorsal-ventral hippocampal axis (A4, Bregma:  $\sim +6.6$ mm). **B**, The immunoreactivity of dendritic marker microtubule-associated protein 2 (MAP2) in the dorsal hippocampal CA1 (B1, Bregma:  $\sim -3.8$ mm) as well as in the ventral level (B2, Bregma:  $\sim +6.6$ mm). **C**, Quantitative analysis of HCN1- and MAP2-immunoreactivity in strata radiatum (SR) and lacunosum-moleculare (SLM). For each area, 12 matched sections from 4 brains (3 sections/area/brain) were analyzed, and optical density was calibrated by subtracting the density of the corpus callosum (white matter background value). Adjacent sections were used for HCN1 and MAP2. Two-way repeated-measures ANOVA was used, followed by Bonferroni's post hoc comparison. \*,  $p < 0.05$ ; \*\*,  $p < 0.01$ . Abbreviations: SP, stratum pyramidale; SR, stratum radiatum; SLM, stratum lacunosum-moleculare; DG, dentate gyrus. Asterisks demarcate the hippocampal fissure.



**Figure 9.**

Application of cAMP increases  $I_h$  amplitude and kinetics, and resonance in ventral somata. **A**, Impedance curve before and after cAMP in a ventral cell. Histograms show the increase of  $F_{res}$  and  $Q$  following cAMP application in ventral, but not dorsal cells. **B**, Example of the effect of cAMP on  $I_h$  current and activation curve in the same cell as in (A). Histograms of  $I_h$  current properties before and after cAMP. Note the increase in amplitude and faster kinetics in ventral, but not dorsal cells. **C**, Temporal summation was decreased by cAMP in ventral cells, but not in dorsal cells. \*,  $p < 0.05$ ; \*\*,  $p < 0.01$ ; \*\*\*,  $p < 0.001$  (paired t-test).

**Table 1**

Morphometric properties of CA1 pyramidal cells from the dorsal and ventral hippocampus.

<i>Cell morphology</i>	Mean cell body area ( $\mu\text{m}^2$ )	Mean total length of apical dendrite ( $\mu\text{m}$ )	Mean diameter of the 1 <sup>st</sup> order apical dendrite ( $\mu\text{m}$ )	Mean total length of basal dendrites ( $\mu\text{m}$ )
<i>Dorsal (n=5)</i>	267 $\pm$ 52	3918 $\pm$ 891	3.48 $\pm$ 0.61	1602 $\pm$ 373
<i>Ventral (n=5)</i>	256 $\pm$ 22	3815 $\pm$ 494	3.34 $\pm$ 0.08	1849 $\pm$ 271

**Table 2**

Recording distance from the soma (for dendritic recordings), basic properties, resonance and  $I_h$  parameters of dendrites and somata in dorsal and ventral cells for whole cell recordings.

	Dorsal dendrite (n=19)	Ventral dendrite (n=15)	Dorsal soma (n=46)	Ventral soma (n=40)
<b>Basic properties</b>				
Distance ( $\mu\text{m}$ )	133 $\pm$ 6	127 $\pm$ 4		
RMP (mV)	-60.1 $\pm$ 0.5	-58.9 $\pm$ 0.6	-58.4 $\pm$ 0.3	-60.3 $\pm$ 0.5 *
Rin (M $\Omega$ )	189 $\pm$ 19	218 $\pm$ 23	100 $\pm$ 8	146 $\pm$ 12 *
C (pF)	95 $\pm$ 8	93 $\pm$ 10	97 $\pm$ 5	104 $\pm$ 6
RS (M $\Omega$ )	31 $\pm$ 2 [20–40]	29 $\pm$ 2 [18–40]	22 $\pm$ 1 [9–39]	19 $\pm$ 1 [8–38]
<b>Resonance</b>				
Fres (Hz)	3.3 $\pm$ 0.2	3.9 $\pm$ 0.4	4.6 $\pm$ 0.1	3.8 $\pm$ 0.2 **
Q	1.15 $\pm$ 0.01	1.18 $\pm$ 0.02	1.27 $\pm$ 0.01	1.16 $\pm$ 0.01 ***
F <sub>0</sub> (Hz)	2.1 $\pm$ 0.2	2.3 $\pm$ 0.2	2.6 $\pm$ 0.1	1.9 $\pm$ 0.1 **
$\Phi_L$ (rad. Hz)	0.06 $\pm$ 0.01	0.07 $\pm$ 0.02	0.11 $\pm$ 0.01	0.04 $\pm$ 0.01 **
<b><math>I_h</math> properties (whole cell)</b>				
At -140 mV (pA)	328 $\pm$ 19	318 $\pm$ 31	308 $\pm$ 14	272 $\pm$ 10 *
At -90 mV (pA)	147 $\pm$ 9	149 $\pm$ 13	138 $\pm$ 6	122 $\pm$ 5 *
V <sub>1/2</sub> (mV)	-89.8 $\pm$ 0.9	-89.9 $\pm$ 0.9	-88.9 $\pm$ 0.5	-91.3 $\pm$ 0.5 *
$\tau_{\text{fast}}$ (ms)	30 $\pm$ 1	28 $\pm$ 1	20 $\pm$ 1	28 $\pm$ 1 *
$\tau_{\text{slow}}$ (ms)	205 $\pm$ 14	199 $\pm$ 14	190 $\pm$ 13	197 $\pm$ 15

\*, p<0.05;

\*\*, p<0.01;

\*\*\*, p<0.001 (ventral vs. dorsal).



**Table 3**

Recording distance from the soma (for dendritic recordings) and  $I_h$  parameters (current density,  $V_{1/2}$ , tau) of dendrites and somata in dorsal and ventral cells for cell attached recordings at PN11-12, as well as for dendrites recorded at PN14-15.

Cell attached	Dorsal dendrite PN11-12 (n=5)	Ventral dendrite PN11-12 (n=6)	Dorsal soma PN11-12 (n=8)	Ventral soma PN11-12 (n=8)	Dorsal dendrite PN14-15 (n=9)	Ventral dendrite PN14-15 (n=7)
Distance ( $\mu$ m)	126 $\pm$ 8	145 $\pm$ 10			228 $\pm$ 10	234 $\pm$ 7
Current density (pA/ $\mu$ m <sup>2</sup> )	4.7 $\pm$ 0.3	4.8 $\pm$ 1.3	3.8 $\pm$ 0.5	1.9 $\pm$ 0.5 **	9.1 $\pm$ 0.8	7.2 $\pm$ 0.7 *
$V_{1/2}$ (mV)	-97.9 $\pm$ 5.6	-101.6 $\pm$ 5.3	-99.2 $\pm$ 1.8	-109.9 $\pm$ 2.7 ***	-101.6 $\pm$ 2.6	-109.9 $\pm$ 2.9 *
$\tau$ (ms)	23.6 $\pm$ 1.8	33.4 $\pm$ 1.4 **	23.6 $\pm$ 1.6	35.5 $\pm$ 2.2 **	21.9 $\pm$ 1.9	28.4 $\pm$ 2.5 *

\* , p<0.05;

\*\* , p<0.01;

\*\*\* , p<0.001 (ventral vs. dorsal).